

Ying Zhu, Cai-Xia Yang and
Zhong-Jian Jia*State Key Laboratory of Applied Organic
Chemistry, Chemistry and Chemical Engineering
College, Lanzhou University, Lanzhou 730000,
People's Republic of ChinaCorrespondence e-mail: jjiazj@lzu.edu.cn

Key indicators

Single-crystal X-ray study
 $T = 293\text{ K}$
Mean $\sigma(\text{C}-\text{C}) = 0.004\text{ \AA}$
 R factor = 0.038
 wR factor = 0.082
Data-to-parameter ratio = 10.3For details of how these key indicators were
automatically derived from the article, see
<http://journals.iucr.org/e>.

A rearranged labdane diterpene glycoside

The title compound, 13(*R*)-9 β -methyl-1(10),14-dienfriedlabda-13-*O*- α -L-2'-acetylrrhamnonopyanoside or 3-methyl-5-(1,2,5,5-tetramethyl-1,2,3,4,4a,5,6,7-octahydronaphthyl)pent-1-en-3-yl 2-*O*-acetylrrhamnonopyanoside, $\text{C}_{28}\text{H}_{46}\text{O}_6$, is a bicyclic diterpenoid glycoside. The rearranged methyl group (from C10 to C9) is in a β orientation and a double bond is formed between C1 and C10 in the rearrangement. The L-rhamnose group on the cyclohexane ring is equatorial and the configuration of the C atom to which the methylene group is attached is *R*. The cyclohexene, cyclohexane and L-rhamnose rings in the molecule adopt half-chair, chair and chair conformations, respectively. In the crystal structure, molecules are linked by $\text{O}-\text{H}\cdots\text{O}$ hydrogen bonds. The L-2'-acetylrrhamnose hydroxy groups serve as hydrogen-bond donors, forming molecular chains along the *b* axis.

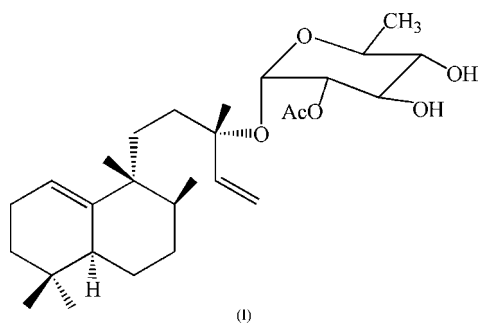
Received 10 January 2006

Accepted 19 January 2006

Online 25 January 2006

Comment

Labdane diterpenoids are among the most common types of diterpenes isolated from terrestrial higher plants and sponges (Hanson, 1997, 1998, 1999, 2000, 2001; Tanaka, *et al.*, 2001). Many of these terpenoids possess significant pharmacological properties, such as cytotoxic, antibacterial, antifungal, anti-inflammatory, analgesic, antitumor and antimutagenic (Ahsan *et al.*, 2003; Itokawa & Morita, 1988; Dimas *et al.*, 1998; Kittakoop *et al.*, 2001; Kubo *et al.*, 2003; Minami *et al.*, 2002; Miyazawa *et al.*, 1995). Therefore, the semisynthesis of minor components from other abundant natural products is of long-standing interest. To date, a number of semisyntheses of these biologically active labdane-type diterpenoids have been reported (Pathak *et al.*, 2005). The current interest of our group in the phytochemical study of northwest Chinese plants aims to find new natural compounds with interesting biological activities. In this connection we have studied labdane diterpenoids (Wang *et al.*, 2002; Yang, *et al.*, 2005).



In this paper, we report the crystal structure and relative stereochemistry of a diterpenoid glycoside, (I), with a rear-

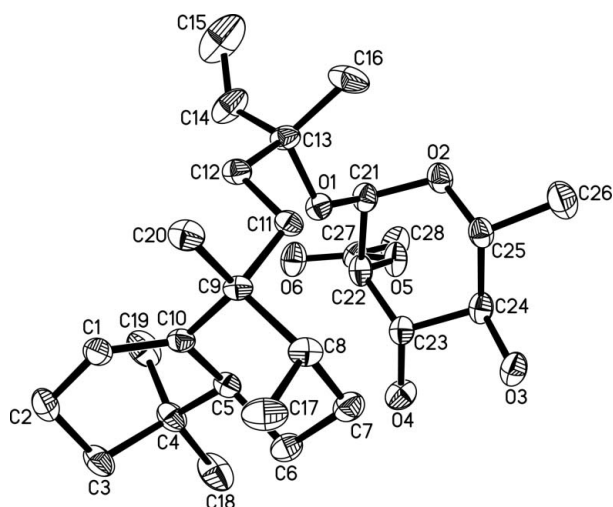


Figure 1
The molecular structure of (I). Displacement ellipsoids are drawn at the 50% probability level. H atoms have been omitted.

ranged labdane skeleton, from *Aster homochlamydeus* Handmazz. Crystal structures of typical labdane-type diterpenoids have previously been reported (Nagashima *et al.*, 1995; Bernardinelli *et al.*, 1988; Tavanaiepour *et al.*, 1987; Bjämer *et al.*, 1968).

Compound (I) has the molecular formula $C_{28}H_{46}O_6$, established by FAB-MS, which gave $[M+Na]$ at m/z 501 and $[M+Li]$ at 485. Its IR spectrum showed the presence of hydroxyl functionalities (3483 and 3373 cm^{-1}). Since compound (I) has five degrees of unsaturation, it must contain one glycoside, two olefin bonds and two carbocyclic rings.

As shown in Fig. 1, compound (I) is a bicyclic diterpene glycoside of a C_{20} skeleton with a terminal double bond, $C14=C15$. It is slightly different from typical labdane diterpenes. The methyl group (C20) normally located at C10 is moved to C9 and the C1–C10 bond length indicates double-bond character. The methyl group at C9 is in the β -orientation. Thus compound (I) is a diterpenoid with a rearranged labdane-type skeleton (Urones *et al.*, 1994; Feresin *et al.*, 2003).

The bond distance between C9 and C20 is $1.532(3)\text{ \AA}$. The olefin bond distance in a carbocyclic ring, between C1 and C10, is $1.328(4)\text{ \AA}$, and corresponding bond angles show the characteristics of an olefin [$C2-C1-C10 = 125.4(3)$ and $C1-C10-C5 = 121.2(3)^\circ$]. The bond distance between C14 and C15 is $1.264(4)\text{ \AA}$, with $C13-C14-C15 = 128.8(5)^\circ$. The structure demonstrates that an equatorial 1,2'-acetylrrhamnose group and a methyl group are attached at C13. The bond lengths to these substituents [$C13-C16 = 1.509(4)\text{ \AA}$ and $C13-O1 = 1.455(3)\text{ \AA}$] are in good agreement with the standard values observed for C–C (methyl) and C–O (sugar hydroxyl) distances, respectively (Allen *et al.*, 1987). The *R* configuration at C13 is confirmed unambiguously (Bernardinelli *et al.*, 1988). Furthermore, the relative configurations at C4, C5, C8 and C9 were also determined, as shown in Fig. 1.

The endocyclic torsion angles (Table 1) show that the cyclohexene ring is in the half-chair conformation (Daux *et al.*, 1974, 1976), which has an approximate twofold axis passing

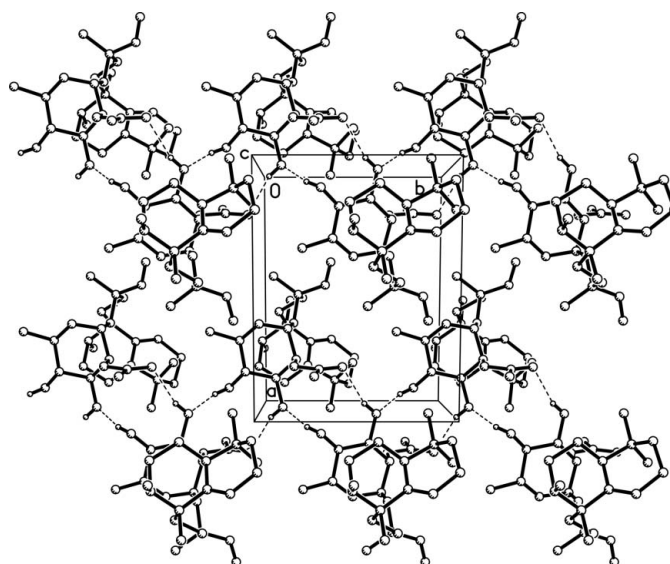


Figure 2
The molecular packing of (I), viewed along the *c* axis. Dashed lines indicate hydrogen bonds. H atoms not involved in hydrogen bonding have been omitted.

through the mid-point of the C3–C4 bond and the mid-point of the C1–C10 bond, with an asymmetry parameter ΔC_2 of 0.88° . The cyclohexane ring and 1,2'-acetylrrhamnose groups are in approximate chair conformations (Daux *et al.*, 1974, 1976); their conformations are similar to others in the literature (Tavanaiepour *et al.*, 1987). The molecules of (I) are associated in the crystal state by O–H...O hydrogen bonds between the hydroxyl functions in the 1,2'-acetylrrhamnose group (Table 2). As shown in Fig. 2, the molecules form columns along the *b* axis.

Experimental

The extraction and isolation of compound (I) have already been described (Yang *et al.*, 2005). Compound (I) was dissolved in chloroform and acetone (1:1 *v/v*), and slow evaporation gave crystals suitable for X-ray diffraction. The optical rotation is $[\alpha]_D^{25} = +33.0^\circ$ (*c* 0.4, $CHCl_3$) and its melting point is 420 K.

Crystal data

$C_{28}H_{46}O_6$
 $M_r = 478.65$
Monoclinic, $P2_1$
 $a = 11.542(2)\text{ \AA}$
 $b = 8.793(2)\text{ \AA}$
 $c = 14.095(2)\text{ \AA}$
 $\beta = 102.14(1)^\circ$
 $V = 1398.6(4)\text{ \AA}^3$
 $Z = 2$

$D_x = 1.137\text{ Mg m}^{-3}$
Mo $K\alpha$ radiation
Cell parameters from 42 reflections
 $\theta = 2.7\text{--}14.3^\circ$
 $\mu = 0.08\text{ mm}^{-1}$
 $T = 293(2)\text{ K}$
Prism, colourless
 $0.58 \times 0.40 \times 0.16\text{ mm}$

Data collection

Siemens P4 diffractometer
 ω scans
Absorption correction: none
3583 measured reflections
3289 independent reflections
1841 reflections with $I > 2\sigma(I)$
 $R_{int} = 0.020$

$\theta_{max} = 27.1^\circ$
 $h = 0 \rightarrow 14$
 $k = 0 \rightarrow 11$
 $l = -18 \rightarrow 17$
3 standard reflections
every 97 reflections
intensity decay: 3.0%

Refinement

Refinement on F^2
 $R[F^2 > 2\sigma(F^2)] = 0.038$
 $wR(F^2) = 0.082$
 $S = 0.81$
 3289 reflections
 320 parameters
 H-atom parameters constrained

$w = 1/[\sigma^2(F_o^2) + (0.04P)^2]$
 where $P = (F_o^2 + 2F_c^2)/3$
 $(\Delta/\sigma)_{\max} < 0.001$
 $\Delta\rho_{\max} = 0.12 \text{ e } \text{Å}^{-3}$
 $\Delta\rho_{\min} = -0.13 \text{ e } \text{Å}^{-3}$
 Extinction correction: *SHELXL97*
 Extinction coefficient: 0.0125 (13)

Table 1

Selected torsion angles ($^\circ$).

C10—C1—C2—C3	15.5 (4)	C4—C5—C10—C1	17.8 (3)
C1—C2—C3—C4	-44.8 (4)	C6—C5—C10—C9	62.0 (3)
C2—C3—C4—C5	60.4 (3)	C8—C9—C10—C5	-59.5 (3)
C3—C4—C5—C10	-45.6 (3)	C25—O2—C21—C22	-59.6 (3)
C10—C5—C6—C7	-55.5 (3)	O2—C21—C22—C23	52.9 (3)
C5—C6—C7—C8	52.5 (3)	C21—C22—C23—C24	-50.6 (3)
C6—C7—C8—C9	-50.3 (3)	C22—C23—C24—C25	52.8 (3)
C7—C8—C9—C10	51.3 (3)	C21—O2—C25—C24	60.2 (3)
C2—C1—C10—C5	-2.1 (4)	C23—C24—C25—O2	-55.0 (3)

Table 2

Hydrogen-bond geometry ($\text{Å}, ^\circ$).

$D-H \cdots A$	$D-H$	$H \cdots A$	$D \cdots A$	$D-H \cdots A$
$O3-H3O \cdots O4^i$	0.82	1.91	2.714 (3)	165
$O4-H4O \cdots O6^i$	0.82	2.17	2.968 (3)	163

Symmetry code: (i) $-x, y - \frac{1}{2}, -z + 1$.

All H atoms were refined as riding on their parent atoms with C—H distances of 0.93–0.98 Å and O—H = 0.82 Å, and with $U_{\text{iso}}(\text{H}) = 1.2U_{\text{eq}}(\text{C}, \text{O})$. In the absence of significant anomalous scattering, Friedel pairs were merged and the absolute configuration is arbitrary.

Data collection: *XSCANS* (Siemens, 1994); cell refinement: *XSCANS*; data reduction: *SHELXTL* (Sheldrick, 1994; program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *SHELXTL*; software used to prepare material for publication: *SHELXTL*.

The authors thank the National Natural Science Foundation of China (grant No. 29972017) for financial support.

References

Ahsan, M., Islam, S. K. N., Gray, A. I. & Stimson, W. H. (2003). *J. Nat. Prod.* **66**, 958–961.
 Allen F. H., Kennard, O., Waston, D. G., Brammer, L., Orpen, A. G. & Taylor, R. (1987). *J. Chem. Soc. Perkin Trans. 2*, pp. S1–19.
 Bernardinelli, G., Vial, C., Starkemann, S. & Näf, F. (1988). *Acta Cryst.* **C44**, 715–717.
 Bjåmer, K., Ferguson, G. & Melville, R. D. (1968). *Acta Cryst.* **B24**, 855–865.
 Daux, W. L., Weeks, C. M. & Rohrer, D. C. (1974). *Topics of Stereochemistry*, Vol. 8, edited by E. L. Eliel & N. L. Allinger, pp. 165–187. New York: John Wiley.
 Daux, W. L., Weeks, C. M. & Rohrer, D. C. (1976). *Topics of Stereochemistry*, Vol. 9, edited by E. L. Eliel & N. L. Allinger, pp. 279–289. New York: John Wiley.
 Dimas, K., Demetzos, C., Marsellos, M., Sotiriadou, R., Malamas, M. & Kokkinopoulos, D. (1998). *Planta Med.* **64**, 208–211.
 Feresin, G. E., Tapia, A., Gimenez, A., Ravelo, A. Z., Sortino, M. & Schmeda-Hirschmann, G. (2003). *J. Ethnopharmacol.* **89**, 71–80.
 Hanson, J. R. (1997). *Nat. Prod. Rep.* **14**, 245–258.
 Hanson, J. R. (1998). *Nat. Prod. Rep.* **15**, 93–106.
 Hanson, J. R. (1999). *Nat. Prod. Rep.* **16**, 209–219.
 Hanson, J. R. (2000). *Nat. Prod. Rep.* **17**, 165–174.
 Hanson, J. R. (2001). *Nat. Prod. Rep.* **18**, 88–94.
 Itokawa, H. & Morita, H. (1988). *Planta Med.* **54**, 117–120.
 Kittakoop, P., Wanasith, S., Watts, P., Kramyu, J., Tanticharoen, M. & Thebtaranonth, Y. (2001). *J. Nat. Prod.* **64**, 385–388.
 Kubo, I., Fujita, K. I., Kubo, A., Nihei, K. I. & Lunde, C. S. (2003). *J. Agric. Food. Chem.* **51**, 3951–3957.
 Minami, T., Wada, S. I., Tokuda, H., Tanabe, G., Muraoka, O. & Tanaka, R. (2002). *J. Nat. Prod.* **65**, 1921–1923.
 Miyazawa, M., Shimamura, H., Nakamura, S. & Kameoka, H. (1995). *J. Agric. Food. Chem.* **43**, 3012–3015.
 Nagashima, F., Tanaka, H., Takaoka, S. & Asakawa, Y. (1995). *Phytochemistry*, **45**, 353–363.
 Pathak, A., Aslaoui, J. & Morin, C. (2005). *J. Org. Chem.* **70**, 4184–4187.
 Sheldrick, G. M. (1994). *SHELXTL*. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.
 Sheldrick, G. M. (1997). *SHELXS97* and *SHELXL97*. University of Göttingen, Germany.
 Siemens (1994). *XSCANS*. Version 2.10b. Siemens Analytical X-ray Instruments Inc., Madison, Wisconsin, USA.
 Tanaka, J., Marriott, G. & Higa, T. (2001). *J. Nat. Prod.* **64**, 1468–1470.
 Tavaniaepour, I., Watson, W. H., Gao, F. & Mabrg, T. J. (1987). *Acta Cryst.* **C43**, 754–756.
 Urones, J. G., Marcos, I. S., Basabe, P., Sexmero, M. J., Carrillo, H. & Melchor, M. J. (1994). *Phytochemistry*, **37**, 1359–1361.
 Wang, W.-S., Li, E.-W. & Jia, Z.-J. (2002). *Pharmazie*, **57**, 343–345.
 Yang, C.-X., Zhang, Q. & Jia, Z.-J. (2005). *Pharmazie*, **60**, 461–463.